

A Novel Direct Route to 2-Deoxy-2-fluoro-aldoses and their Corresponding Derivatives

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Abstract: A new type of reactive *N*-(2-deoxy-2-fluoro-glycosyl) compound is formed by regioselective *syn*-addition of the electrophilic *N*-F reagent SelectfluorTM to glycals. By subsequent treatment with nucleophiles, 2-deoxy-2-fluoro-aldoses and various C-1-substituted derivatives thereof are easily accessible. Furthermore, the reaction with D-galactal and D-arabinal proceeds stereoselectively, thus allowing the synthesis of 2-deoxy-2-fluoro-D-galactose- and -D-arabinose-derivatives in multi-gram quantities. © 1998 Elsevier Science Ltd. All rights reserved.

INTRODUCTION

Compounds containing 2-deoxy-2-fluoro-aldoses are of increasing importance in biochemical studies¹ as well as medicinal chemistry research.² Since their chemical synthesis is hampered by a broad range of restrictions, a vast number of approaches is known.³ These include strategies like electrophilic addition of *O*-F reagents (*e.g.* acetyl hypofluorite) to glycals.⁴ In analogy to this method, a program to investigate the scope and limitations of applying the new class of electrophilic *N*-F fluorinating agents⁵ for the transformation of glycals⁶ into 2-deoxy-2-fluoro-sugars was undertaken[‡]. From the list of commercially available reagents, 1-fluoro-pyridinium tetrafluoroborate (**1a**), 1-fluoro-2,**4**,6-trimethylpyridinium triflate (**1b**), *N*-fluorobenzene-sulfonimide (**2**) and 1-chloromethyl-4-fluoro-1,4-diazonia-bicyclo[2.2.2]octane bis(tetrafluoroborate)⁷ (SelectfluorTM, F-TEDA-BF₄, **3**) were selected (Fig.1).

Figure 1. Structures of N-F reagents tested

RESULTS AND DISCUSSION

Treatment of 3,4,6-tri-O-acetyl-1,5-anhydro-2-deoxy-D-lyxo-hex-1-enitol (3,4,6-tri-O-acetyl-D-galactal, 4) with 1a or 1b, in acetonitrile and temperatures up to 80 °C, did not lead to a reaction. Employing compound 2, after 24 hours at 80 °C, a single addition product, N,N-di(phenylsulfonyl)-3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-β-D-galactopyranosylamine (5), was obtained. Application of 3, at room temperature, in various solvents and their mixtures with methanol and water, respectively, caused (as evidenced by t.l.c.- and ¹⁹F NMR monitoring) rapid and complete consumption of 4 with the simultaneous formation of a range of fluorinated carbohydrates (Scheme 1).

NP = Nitrophenyl, DNP = Dinitrophenyl **Scheme 1.**

Isolation of the products formed using precipitative and chromatographic methods, followed by elucidation of the individual structures by means of NMR spectroscopy⁸ and correlation (Table 1) with their respective signals in the ¹⁹F NMR spectra⁹ allowed the following conclusions.

Introduction of fluorine occurs regio- and stereospecifically with the sole formation of 2-deoxy-2-fluoro-D-galactopyranose derivatives.

Among the products formed, a new N-glycosyl compound, 1-(3,4,6-tri-O-acetyl-2-deoxy-2-fluoro- α -D-galactopyranosyl)-4-chloromethyl-1,4-diazonia-bicyclo[2.2.2]octane bis(tetrafluoroborate) (6), predominates.

Anomeric mixtures of (OH-1-unprotected) 3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-D-galactopyranose (7a) or the respective methyl glycosides (7b) are obtained in the presence of water or methanol.

In the absence of external nucleophiles, participation of fluoride ion (liberated from tetrafluoroborate) and the solvent (acetonitrile or N,N-dimethylformamide) gives rise to 3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-O-galactopyranosyl fluoride (7c), anomeric mixtures of N-acetyl-3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-D-galactopyranosylamine (7d) or 3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-1-O-formyl-D-galactopyranose (7e).

| Table 1. Product Ratio | [%] | as Determined by | ^{i 19} F NMR | Spectroscopy ⁹ |
|------------------------|-----|------------------|-----------------------|---------------------------|
|------------------------|-----|------------------|-----------------------|---------------------------|

| solvent | 6 | 7a | 7b | 7e | 7d | 7e | by-products [a] |
|---|----|----|----|----|----|----|-----------------|
| CH ₃ NO ₂ | 73 | | | 23 | | | 4 |
| CH ₃ NO ₂ /D ₂ O (5:1) | 58 | 30 | | | | | 12 |
| CH ₃ NO ₂ /CD ₃ OD (5:1) | 49 | | 45 | 4 | | | 2 |
| CH ₃ CN | 54 | | | 14 | 27 | | 5 |
| CH ₃ CN/D ₂ O (5:1) | 45 | 23 | | | 30 | | 2 |
| Acetone/D ₂ O (5:1) [b] | 49 | 49 | | | | | 2 |
| DMF/D ₂ O (5:1) | 46 | 31 | | | | 22 | 1 |

[a] summation of unidentified fluorinated carbohydrates [b] additionally, formation of fluoro acetone $[\delta$ -226.9 ppm, J 47 and 47 Hz] occurred

Since reaction monitoring after total consumption of 4 had revealed only slow changes within the proportions of fluorinated products, the mixtures containing water or methanol were heated. Within a few hours, quantitative substitution of the ammonium group at C-1 of compound 6 by the protic co-solvent, to form respectively 7a and 7b, occurred. With neat aprotic solvents (besides increased formation of 7c) no other clean reaction was observed. Intermediate 6 was isolated by precipitation. Subsequent treatment with sodium azide, 4-nitrophenolate or 2,4-dinitrophenolate well bromide, potassium as magnesium bis(trimethylsilyl)thymine, in acetonitrile or nitromethane, gave 3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-β-Dgalactopyranosyl azide (8a), the corresponding α -bromide (8d), 4-nitrophenyl (8b) or 2,4-dinitrophenyl β glycoside (8c) and 1-(3,4,6-tri-O-acetyl-2-deoxy-2-fluoro-β-D-galactopyranosyl)thymine (8e), respectively, in good yields (Table 2). Interestingly, treatment of 4 with 3 in the presence of bromide or phenolate ions did not yield fluorinated carbohydrates. Intermediate 6, with thiophenolate or xanthate ions, underwent reaction 11 other than nucleophilic substitution at C-1 of the carbohydrate moiety. When performing the reaction of 4 with 3 in boiling acetonitrile, besides 7c, 4,6-di-O-acetyl-1,5-anhydro-2-deoxy-D-threo-hex-1-en-3-ulose 12 (9) was also isolated from the orange coloured reaction mixture.

Table 2. Selected Transformations

| substrate | No | product | No | pro- cedure | yield [%] | [\alpha] _D (c, solvent) mp. R _f [a] |
|-----------------|----|--------------------|-----|----------------|--------------|---|
| AcO OAc | 4 | Aco O N BF4 | 6 | A | 82 [b] | +10.8° (2.0, MeCN) (foam) 0.0 |
| | | AcO WOH | 7a | В1 | 79 | [c] (-) syrup 0.42 |
| | | Aco Aco OAc | 7b | B1 | 76 | [c] (-) syrup 0.61 |
| | | AcO F | 7c | В3 | 52 | +103° (0.9, CHCl ₃) 58°C 0.55 |
| AcO OAC BF. BF. | 6 | Aco N ₃ | 8a | С | 62 | +89.5° (3.0, CHCl ₃) 84°C 0.63 |
| | | AcO ODNP | 8c | С | 61 | -3.6° (1.0, CHCl ₃) 160°C 0.45 |
| | | AcO F Br | 8d | C | 66 | +162.0° (0.8, CHCl ₃) syrup 0.63 |
| • | | AcO Thymine | 8e | С | 63 | +18.6° (0.9, CHCl ₃) syrup 0.21 |
| AcO AcO | 10 | OAc OAc | 12d | B2 | 68 [d] | [c] (-) syrup 0.57 -187.5° |
| | | OAC P | 12b | В3 | 45 | (1.1, CHCl ₃) 64°C 0.65 |

[a] ethyl acetate/cyclohexane (1:1) [b] crude yield [c] mixture of anomers [d] additionally 7% of the corresponding D-ribo-configurated isomers were obtained (α/β 1:2)

Similar results to those found with D-galactal 4 were obtained from the reaction of 3,4-di-*O*-acetyl-1,5-anhydro-2-deoxy-D-*erythro*-pent-1-enitol (di-*O*-acetyl-D-arabinal, 10) with 3 (Scheme 2). When carried out in acetone/water, exclusive formation of products with D-*arabino*-configuration, namely 1-(3,4-di-*O*-acetyl-2-deoxy-2-fluoro-β-D-arabinopyranosyl)-4-chloromethyl-1,4-diazonia-bicyclo[2.2.2]octane bis(tetrafluoroborate) (11¹³) and the anomers of 3,4-di-*O*-acetyl-2-deoxy-2-fluoro-D-arabinopyranose¹⁴ (12a), was observed. The same experiment performed in nitromethane/water produced a minor proportion of the D-*ribo*-isomer. Reaction in acetonitrile led to β-configurated difluoride 12b as well as, through participation of the solvent, *N*-acetyl-3,4-di-*O*-acetyl-2-deoxy-2-fluoro-D-arabinopyranosylamine 16 (12c).

In contrast to the results obtained with D-galactal 4, pronounced stereoselectivity was not observed in the reaction of derivatives of 1,5-anhydro-2-deoxy-D-*arabino*-hex-1-enitol (D-glucal) including disaccharidic analogues derived from maltose, cellobiose and lactose, respectively. Here, without exception, mixtures of the corresponding derivatives of 2-deoxy-2-fluoro-D-glucopyranose and -D-mannopyranose [including (2-deoxy-2-fluoro-glycosyl) ammonium intermediates¹⁷ of type 6 (*gluco*) and 11 (*manno*)] were formed. The composition of these mixtures also varied depending on the nature of the protecting group. When starting from *O*-unprotected D-glucal, an inseparable mixture of 1,6-anhydro-2-deoxy-2-fluoro-β-D-gluco- and -manno pyranose¹⁹ was obtained in moderate yield.

Concerning the mechanism of this "glycal-SelectfluorTM reaction", conclusions can be drawn from the unique stereochemistry of the direct addition products (6 or 11). In each case, even under inversion of otherwise preponderating chair conformations, they possess axially oriented fluorine and equatorially arranged ammonium substituents. For their formation, a concerted *syn*-addition, not yet observed with *N*-halogeno compounds²⁰ including *N*-F reagents, may be anticipated. In addition, depending on the reaction temperature, at least two other competing pathways²¹ appear to exist. Possibly, formation of a 2-deoxy-2-fluoro-glycosyl species (*e.g.* glycosylium ion) occurs, which spontaneously reacts with nucleophiles to give compounds of the type 7a-e (thus diminishing the attainable yield of the products 8a-e). Additionally, at higher temperatures an obviously radical initiated oxidation process with formation of compounds like 4,6-di-*O*-acetyl-1,5-anhydro-2-deoxy-D-*threo*-hex-1-en-3-ulose (9) takes place. Whereas for the reaction of intermediate 6, with nucleophiles, to yield 1.2-*trans*-configurated products 8a, b, c and e, a S_N2-mechanism is obvious, compounds such as 7c and 8d must be formed through a subsequent anomerization to the thermodynamically more stable anomer.

From these findings, short and simple procedures for the preparation of derivatives of 2-deoxy-2-fluoro-D-galacto- and -D-arabinopyranose, valuable in biochemical studies as well as chemical synthesis, were established; their NMR data are collected in Tab. 3 (¹³C), Tab. 4 (¹H) and Tab. 5 (¹⁹F), respectively.

Table 3. 13 C NMR Data (CDCl₃), δ [ppm] (J [Hz])

| No | δ C-1 | δ C-2 | δ C-3 | δ C -4 | δ C-5 | δ C-6 | substituents [a] |
|--------------|---------------------------|--------------------------|------------------------|------------------------|--------------------------|-------|---------------------------------|
| | $(J_{\text{C1-F2}})$ | $(J_{	ext{C2-F2}})$ | $(J_{\mathrm{C3-F2}})$ | $(J_{\mathrm{C4-F2}})$ | | | |
| 5 | 86.6 | 86.0 | 72.2 | 67.5 | 73.7 | 60.7 | PhSO ₂ 141.0/ 139.7/ |
| | (26.1) | (186.1) | (18.6) | (8.2) | | | 134.2/ 134.0/ 129.0/ |
| | | | | | | | 128.8/ 128.6 |
| 6 | 88.7 | 84.8 | 66.8 | 63.9 | 75.9 | 59.4 | TEDA 50.8/ 50.2 |
| | (15.0) | (188.1) | (28.8) | (-) | | | CH ₂ Cl 69 .6 |
| $7a\alpha$ | 90.7 | 85.8 | 68.0 | 68.8 | 66.3 | 61.7 | |
| | (21.4) | (189.1) | (18.8) | (7.8) | | | |
| 7a β | 94.9 | 89.1 | 71.1 | 67.9 | 70.8 | 61.5 | |
| | (23.1) | (186.3) | (19.0) | (9) | | | |
| $7b\alpha$ | 97.5 | 85.5 | n.r. | 68.7 | 66.4 | 61.6 | OMe 55.7 |
| | (20) | (185) | | (7.5) | | | |
| 7 b β | 101.7 | 88.0 | 71.1 | 67.7 | 70.7 | 61.1 | OMe 57.4 |
| | (22.8) | (186.9) | (18.9) | (8.4) | | | |
| 7c | 104.1 | 84.4 | 67.7 | 67.8 | 69.0 | 61.1 | |
| | (23.5/ | (191.2/ | (25.1) | (1.6) | | | |
| | $J_{\text{C1-F1}}$ 230.5) | $J_{\text{C2-F1}}$ 25.3) | | | $(J_{C5-F1} 3.2)$ | | |
| 7d β | 77.7 | 86.9 | 71.6 | 68.0 | 72.5 | 61.1 | NHAc 169.9/23.4 |
| | (23.3) | (188.2) | (18.7) | (8.1) | | | |
| 7eα | 88.8 | 84.1 | 67.6 | 68.1 | 69.1 | 61.1 | OCHO 158.5 |
| | (24) | (191.7) | (16) | (9.5) | | | |
| 7 e β | 91.1 | 86.6 | 70.8 | 67. 4 | 72.0 | 60.9 | OCHO 158.5 |
| | (24.9) | (188.7) | (18.7) | (8.2) | | | |
| 8a | 88.0 | 87.4 | 70.9 | 67.5 | 72.8 | 61.1 | |
| | (23.1) | (188.7) | (18.8) | (8.2) | | | |
| 8b | 98.2 | 87.3 | 70.8 | 67.4 | 71.5 | 61.3 | <i>p</i> -NP 116.8 125.8 |
| | (23.8) | (189.2) | (19.1) | (8.2) | | | 143.4 161.1 |
| 8c | 98.9 | 87.0 | 70.4 | 67.0 | 72.0 | 61.2 | DNP 117.7/ 121.8/ |
| | (24.7) | (190.0) | (19.2) | (8.2) | | | 128.7/ 140.0/ 142.2/ |
| | | | | | | | 153.6 |
| 8d | 86.9 | 84.2 | 68.9 | 67.5 | 71.3 | 60.7 | |
| | (25.6) | (195.0) | (18.0) | (7.6) | | | |
| 8e | 80.2 | 85.8 | 71.2 | 67.9 | 73.9 | 61.2 | Thymine 112.7/ |
| | (24.6) | (190.1) | (18.3) | (8.0) | | | 133.9/ 150.6/ 163.37/ |
| | | | | | | | 12.7 |
| 12b | 104.7 | 85.0 | 67.3 | 68.8 | 62.6 | - | |
| | (23.3/ | (190.8/ | (18.9) | (7.7) | | | |
| | $J_{\text{C1-F1}}$ 229.5) | $J_{\text{C2-F1}}$ 25.6) | | | $(J_{\text{C5-F1}} 3.3)$ | | |

[a] data for O-acetyl groups are omitted

Table 4. ¹H NMR Data (CDCl₃), δ [ppm] (J [Hz])

| No | δ H-1 | δ H-2 | δ H-3 | δ H-4 | δ H-5 | δ H-6a | δ H-6b | substituents [a] |
|--------------|--------------------------|-------------------------------|----------------------|----------------------|---------------------------|--------------------|----------------------|-----------------------------|
| | $(J_{\mathrm{H1-F}},$ | $(J_{ m H2-F},$ | $(J_{\text{H3-F}},$ | $(J_{	ext{H4-F}},$ | $(J_{ m II5-H6a},$ | $(J_{ m H6a-H6b})$ | | |
| | $J_{ m H1-H2})$ | J _{H2-H3} ,) | $J_{\mathrm{H3-H4}}$ | J _{H4-H5}) | J _{H5-H6b}) | | | |
| 5 | 5.62 | 5.66 | 5.16 | 5.43 | 3.88-4.06 | | PhSO ₂ | |
| | (2.8/9.0) | (53.1/9.2) | (13.7/3.3) | (-,2.6) | n.r. [b] | | 7.5-7.7, 8.0-8.2, | |
| | | | | | | 4.0 | 40.40 | (m) |
| 6 | | 5.2 | | | | -4.8 | 4.0-4.3 | TEDA 4.0-4.3, |
| | | n | .r. | | n | ı.r. | n.r. | (m) CH ₂ Cl 5.27 |
| | 5 20 5 54 | 4.77.5 | 5.20 | <i>5.5.</i> 4 | 4.40 | 2.05 | 4.15 | (s) |
| 7 a α | 5.38-5.54, | 4.75 | 5.38- | | 4.48 | 3.95- | | |
| | n.r. | (50.2/9.7) | n | .r. | (6.6/6.6) | n. | r. | |
| 7 0 | (-,3.7) | 4 47 | 5 1 1 | | 2.05 | 4 15 | | |
| 7a β | 5.38-5.54, | 4.47 | 5.11 | | 3.95-4 | | | |
| | n.r. | (51.3/9.8) | (12.9/3.5) | | n.ı | r. | | |
| 71 | (n.r./7.5) | 4.75 | | | | | | OCH ₃ |
| 7bα | 5.0 | 4.75 | | | n.r. | | | 3.46 (s) |
| 71.0 | (-/ 3.5) 4.49 | (50/10) 4.48 | 5.10 | 5.41 | 3.92 | 4.05- | 1 25 | OCH ₃ |
| 7b β | 4.49 (2.2/ 7.7) | | | | (6.6/6.6) | 4.05-4 n.i | | 3.59 (s) |
| 7.0 | 5.84 | (52.7/ 9.6) 4.77 | (13.4/ 3.5) 5.40 | (n.r.) 5.53 | 4.40 | 4.0- | | 3.39 (3) |
| 7c | 3.84 (-/ 2.9/ | (48.8/10.1 | (11.1/3.3) | (3.3/1.1) | (6.2/6.2) | n.i | | |
| | • | * | (11.1/ 3.3) | (3.3/ 1.1) | (0.2/ 0.2) | 11.1 | • | |
| 740 | $J_{\text{HI-FI}}$ 53.1) | $J_{\text{H2-F1}}$ 23.3) 4.48 | 5.17 | 5.44 | | 4.0-4.2 | | NH 6.48 |
| 7d β | 5.35 (3.0/ 9.0) | (51.4/ 9.5) | (12.4/3.5) | | | n.r. | | $(J_{\text{NH-H}1} 9.4)$ |
| 700 | 6.56 | 4.92 | 5.3- | | | 4.0-4.4 | | OCHO |
| 7eα | (-/ 3.8) | (50/10) | 5.5 n. | | | n.r. | | 8.16 (s) |
| 7e β | 5.88 | 4.66 | 5.19 | 5.52 | | 4.0-4.4 | | OCHO |
| /ep | (4.1/8.0) | (51.3/ 9.8) | (13.1/3.5) | (3.5/1.8) | | n.r. | | 8.12 (s) |
| 8a | 4.81 | 4.42 | 5.12 | 5.42 | | 3.88-4.18 | | 0.12 (5) |
| Оа | (4.0/8.5) | (50.8/9.7) | (13.0/3.5) | (2.6/1) | | n.r. | | |
| 8b | 5.29 | 4.81 | 5.25 | 5.50 | | 4.05-4.30 | | <i>p</i> -NP 7.13 /8.20 |
| 00 | (4.0/7.5) | (51.1/9.8) | (13.3/3.5) | (3/ -) | | n.r. | | (d each) |
| 8c | 5.41 | 4.89 | 5.25 | 5.41 | | 4.05-4.25 | | DNP 7.44 (d) |
| OC | (4.6/7.3) | (50.7/ 9.7) | (13.4/3.5) | (2.5/-) | | n.r. | | 8.42 (dd) |
| | (4.0/ 7.3) | (30.11 7.1) | (15.17 5.5) | (2.3,) | | •••• | | 8.76 (d) |
| 8d | 6.60 | 4.74 | 5.45 | 5.50 | 4.49 | 4.15 | 4.09 | |
| ou | (-/ 4.2) | (50.0/9.6) | (13.4/3.3) | (3.5/1.2) | (6.2/6.8) | (11.4) | | |
| 8e | 5.94 | 4.68 | 5.29 | 5.48 | (3.2. 3.3) | 4.0-4.2 | | Thymine 7.05/ |
| 30 | (3.7/9.2) | (51.3/ 9.6) | (12.3/3.5) | (3.0/-) | | n.r. | | 1.93 (s each) |
| 12b | 5.82 | 4.79 | | 5.45 | 5a: 4.1 | | o: 3.77 | ` / |
| 1 MI 1.7 | (-/ 2.7/ | (48.5/ 10/ | | .r. | (-/-/ | | _{b-H4} 1.6/ | |
| | | $J_{\text{H2-F1}}$ 23.3) | | | $J_{\mathrm{H5a-H5b}}$ 13 | | _{b-F1} 1.6) | |
| | - m-r1 55.1) | - nz-r1 ====) | | | 1454-1150 | , 110 | · · · · · / | |

[[]a] data for acetyl groups are omitted [b] not resolved

| No | δ F-1 | δ F-2 | $J_{ m F2-H1}$ | J _{F2-H2} | $J_{ m F2-H3}$ | J _{F2-H4} | $J_{ m FI-HI}$ | $J_{ m F1-H2}$ | $J_{\mathrm{F1-F2}}$ |
|--------------|--------|--------|----------------|--------------------|----------------|--------------------|----------------|----------------|----------------------|
| 5 | | -203.8 | | 53.3 | 13.5 | | | | |
| 6 [a] | | -204.2 | 25.5 | 46.0 | 10.5 | | | | |
| $7a\alpha$ | | -207.0 | | 50.1 | 10.9 | 3.6 | | | |
| 7aβ | | -207.5 | 3.3 | 51.5 | 13.0 | | | | |
| 7b α | | -209.2 | | 49.8 | 10.9 | 3.4 | | | |
| 7b β | | -207.3 | 2.4 | 52.7 | 13.3 | 2.4 | | | |
| 7c | -152.6 | -211.3 | | 49 | 14 | | 51.1 | 24.1 | 18 |
| 7d β | | -204.9 | | 51.1 | 13.4 | | | | |
| $7e\alpha$ | | -209.5 | | 49.0 | 11,1 | 3.5 | | | |
| 7e β | | -208.6 | 3.5 | 52.9 | 13.0 | 3.0 | | | |
| 8a | | -205.5 | 3.3 | 51.0 | 12.9 | 3.3 | | | |
| 8b | | -207.1 | 3.5 | 51.3 | 13.3 | 3.2 | | | |
| 8c | | -206.7 | 3 | 50 | 15 | 3 | | | |
| 8d | | -195.3 | | 50.9 | 10.4 | 3.3 | | | |
| 8e | | -207.3 | | 51.2 | 12.3 | | | | |
| 12b | -155.2 | -209.3 | | 50 | 14 | | 52 | 22 | 18 |

Table 5. ¹⁹F NMR Data (CDCl₃), δ [ppm], J [Hz]

[a] BF_4 δ -150.2 ppm

EXPERIMENTAL SECTION

General Methods. Melting points were determined with a Tottoli apparatus (Büchi 300) and are uncorrected. Optical rotations were measured with a Jasco DIP-360 digital polarimeter at 589 nm at ambient temperature. NMR spectra were recorded at 300.13 or 200 MHz (¹H), 75.47 or 50.29 MHz (¹³C) and 282.4 MHz (¹⁹F) - using a Bruker MSL 300 and a Varian Gemini 200 apparatus, respectively; as reference standards tetramethylsilane (¹H and ¹³C NMR) and trichlorofluoromethane (¹⁹F NMR) were used. TLC was performed on silica-gel 60 F₂₅₄ precoated aluminum plates (Merck 5554) with detection by charring after spraying with vanillin/sulfuric acid (1%). The R_f values given were determined using ethyl acetate/cyclohexane 1:1 (v:v). For column chromatography, silica gel 60, 230-400 mesh (Merck 9385), was used. "Normal work-up" means: filtration (where heterogenous), evaporation of the solvent *in vacuo*, dissolution of the residue in dichloromethane followed by extraction with water (or 5% sodium hydrogen carbonate and water), drying of the organic phase with sodium sulphate and removal of dichloromethane by distillation.

Samples for ¹⁹F NMR reaction monitoring were taken from solutions (2 ml) of 4 or 10 (40 mg) and 3 (60 mg, 1.2 equivalents) in the mixture of the respective solvent with D₂O or CD₃OD; when neat nitromethane or acetonitrile was used, CDCl₃ was added prior to measurement.

Procedure A: To a solution of 4 (2.00 g, 7.35 mmol) in nitromethane abs. (30 ml) is added 3 (2.60 g, 7.35 mmol) and stirred vigorously at room temperature overnight. After addition of ethyl acetate (40 ml) a precipitate forms (reagent after fluorine delivery), which is removed by filtration and washed with nitromethane (10 ml). Filtrate and washings are concentrated *in vacuo* and the residue is dissolved in acetonitrile (5 ml). Slow addition of ethyl acetate/cyclohexane [100 ml, 1:1 (v:v)] causes deposition of product 6 as an oil, which is separated by centrifugation. After decanting and evaporation of the remaining solvent, crude 6 (3.76 g, 82%) is obtained as a white foam. $R_f(4)$ 0.53; $R_f(6)$ 0.0.

Procedure B1: To a 10% solution of glycal 4 or 10 in nitromethane/water or nitromethane/methanol (5:1, v/v) are added 1.2 equivalents of reagent 3 under vigorous stirring at room temperature. After quantitative consumption of the starting material (a few hours) the mixture is heated to reflux for a period of half an hour.

After evaporation of the solvent, products are isolated by chromatography (ethyl acetate/cyclohexane 1:3). R_f (7a) 0.42; R_f (7b) 0.61; R_f (10) 0.75; R_f (12a) 0.45.

Procedure B2: The residue obtained by procedure B1 is acetylated according to a standard protocol and the peracetates isolated by chromatography (ethyl acetate/cyclohexane 1:4).

Procedure B3: This procedure is identical to B1, except that nitromethane abs. is used as sole solvent. R_f (7c) 0.55; R_f (12b) 0.65.

Procedure C: To a 5% solution of crude 6 in acetonitrile abs. or nitromethane abs. 2 equivalents of the appropriate nucleophile [sodium azide, potassium 4-nitrophenolate, sodium 2,4-dinitrophenolate, magnesium bromide or 2,4-bis(trimethylsilyl)thymine] are added. This mixture is allowed to react, at reflux temperature, until total consumption of 6 (10-30 min). After normal work-up, the products are isolated by chromatography [ethyl acetate/cyclohexane 1:4 (v:v)]; 8c crystallizes from ethanol. R_f (8a) 0.63; R_f (8b) 0.59; R_f (8c) 0.45; R_f (8d) 0.63; R_f (8e) 0.21.

N-Acetyl-3,4,6-tri-*O*-acetyl-2-deoxy-2-fluoro-D-galactopyranosylamine (7d) is obtained from a reaction of 4 (1.00 g, 3.67 mmol) and 3 (1.55 g, 4.38 mmol) in acetonitrile abs. (12 ml). After 15 h at room temperature, 7d¹⁰ (α/β 1:11; 0.15 g, 12%) and 7c (0.40 g, 35%), are isolated by chromatography. R_f (7d) 0.22.

3,4,6-Tri-O-acetyl-2-deoxy-2-fluoro-1-O-formyl-D-galactopyranose (7e, α/β 4:13; 0.18 g, 14%) is isolated from an analogous protocol as described for 7d, but using N,N-dimethylformamide/water (5:1) as solvent. R_f (7e) 0.53.

4,6-Di-O-acetyl-1,5-anhydro-2-deoxy-D-*threo***-hex-1-en-3-ulose (9):** To a boiling solution of **4** (1.00 g, 3.67 mmol) in acetonitrile abs. is added **3** (1.55 g, 4.38 mmol). After quantitative consumption of **4** (ca. 30 min.), the orange coloured reaction mixture is evaporated. From the residue, UV-active product 9^{12} (0.11 g, 14%) as well as 7a, 7c and 7d are isolated by chromatography [ethyl acetate/cyclohexane 1:3 (v:v)]. $R_f(9)$ 0.41.

N,*N*-Di(phenylsulphonyl)-3,4,6-tri-*O*-acetyl-2-deoxy-2-fluoro-β-D-galactopyranosylamine (5). A solution of 4 (1.00 g, 3.67 mmol) and *N*-fluorobenzenesulfonimide (2, 1.39 g, 4.41 mmol) in acetonitrile abs. (15 ml) is heated to 80°C for 24 h. After normal work-up, compound 5 (0.65 g, 30%) is isolated by chromatography [ethyl acetate/cyclohexane 1:3 (v:v)] and recrystallized from ethanol. R_f (5) 0.51; mp. 189 °C; $[\alpha]_D^{20}$ +23.9° (c 1.1, CHCl₃).

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- 9. Due to solvent dependance of the chemical shift, representative values for F-2 signals are given:
 a) MeCN/D₂O: 6 (-203.2 ppm); 7a (-205.7 and 205.9 ppm); 7d (-204.8 and-205.0 ppm);
 b) MeNO₂/CD₃OD: 6 (-203.1 ppm); 7b (-206.3 and -207.9 ppm);
 c) MeNO₂: 6 (-205.1 ppm); 7c (-212.2 ppm);
 d) DMF/ D₂O: 6 (-204.3 ppm); 7a (-205.6 and 205.9 ppm); 7e (-208.1 and -208.8 ppm).
 Characteristic sets of coupling-constants are: 46/25/10 Hz (6); 50/10/3 Hz (α-anomer of 7a, b, d and e);
 52/13/4 Hz (β-anomer of 7a, b, d and e) and 50/20/12 Hz (7e); J < 8 Hz in most cases not resolved.
- 10. The structure of the β -anomer was verified by independent synthesis starting from **8a** by means of reduction followed by N-acetylation.
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- 13. 19 F NMR data: δ -206.6 ppm (*J* 47 and 25 Hz) -150.2 ppm (BF₄); 4 C₁-conformer.
- 14. ¹⁹F NMR data: δ -204.6 ppm (*J* 49 and 8 *Hz*); δ -204.1 ppm (*J* 51 and 12 *Hz*); both anomers in "normal" ₄C¹-conformation. *O*-Acetylation gave 1,3,4-tri-*O*-acetyl-2-deoxy-2-fluoro-D-arabinopyranose (**12d**); for data see: M. Bols, I. Lundt, *Acta Chem. Scand.* **1990**, 44, 252-256.
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- 17. ¹⁹F NMR data and relative proportions (%) for products formed from the reaction of tri-O-acetyl-D-glucal in Me₂CO/D₂O: δ –199.0 ppm (J 50 and I3 Hz) and -199.8 ppm (J 49 and I1 Hz); 2-F-Glc-OH, 29.3%. δ -203.5 ppm (J 50 and 30 Hz) and -221.6 ppm (J 50, 30 and 20 Hz); 2-F-Man-OH; 42.7%. δ -201.3 ppm (J 48, 24 and 10 Hz); ${}_{4}C^{1}$ -conformer of the (2-F-Glc)-intermediate; 14.9%. δ -213.7 ppm (J 50, 30 and 20 Hz); ${}_{4}^{4}C_{1}$ -conformer of the (2-F-Man)-intermediate; 13.2%.
- 18. These results will be published elsewhere.
- 19. ¹⁹F NMR data: δ -188.1 ppm (*J* 45 and 18 Hz) and -207.8 ppm (*J* 45 Hz); rel. intensities 1:3.
- a) N-Iodo- and N-bromopyridinium compounds add to glycals under formation of products with 1,2-trans-diaxial arranged substituents (Lemieux, R. U.; Morgan, A. R. Can. J. Chem. 1965, 43, 2205-2213); see also "halogenoglycosyloxylation" (e.g. Thiem, J.; Klaffke, W. J. Org. Chem. 1988, 54, 2006-2009) and "halogenosulphamidation/sulphamidoglycosyloxylation" (Griffith, D. A.; Danishefsky, S. J. J. Am. Chem. Soc. 1990, 112, 5811-5819); b) for the reaction with N-fluoropyridinium compounds see ref. 6; c) product 5 from these experiments shows 1,2-trans-diequatorial grouping.
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- 22. Syrupy 8b, containing minor impurities, was obtained in 51% yield.